

## Gastrointestinal Health Parasites of Domestic Dogs in Jos North, Plateau State Nigeria: A Fecal Examination Study

Karaye GP<sup>1\*</sup>, Kaze PD<sup>1</sup>, Akinsola OM<sup>2</sup>, Wamtas FI<sup>1</sup>, Kogi AC<sup>1</sup> and Karaye KK<sup>3</sup>

<sup>1</sup>Department of Veterinary Parasitology and Entomology, University of Jos, Plateau State, Nigeria

<sup>2</sup>Department of Theriogenology and Production, University of Jos, Plateau State, Nigeria

<sup>3</sup>Central Diagnostic Laboratory, National Veterinary Research Institute, Vom, Nigeria

Received May 05, 2020; Revised May 22, 2020; Accepted May 24, 2020

### ABSTRACT

Dogs are the most common pet animals worldwide. They may harbor Toxocariasis which remains the most important parasitic infection affecting dogs and pose a risk to animal and human health. There is inadequate information on the risk of this infection on dogs in Jos, Jos North LGA of Plateau State, Nigeria. Thus, a baseline study was undertaken from January 2019 to March 2020 to determine the prevalence of zoonotic gastrointestinal parasite of domestic dogs brought for routine clinical treatment in the Jos University Veterinary Teaching Hospital, Jos Plateau State. A total of 321 fecal samples collected randomly from dogs were processed and examined for helminthes eggs using modified Kato-Katz technique. The overall prevalence of gastrointestinal helminthes was 50.01%. *Toxocara canis* was the most frequently observed gastrointestinal helminthes parasites with a prevalence of 30.63%, while prevalence of 9.68% was recorded for a mixed infection of *Toxocara canis* and *Ancylostoma caninum*, 1.88% was obtained for *Diphylidium caninum* and 7.81% for the single infection of *Ancylostoma caninum* respectively. During the study period, the occurrence of gastrointestinal helminthes based on monthly distribution were; January recorded the highest Prevalence of 35.48% mixed infection (polyparasitism) followed by February (20.00%) in the single infection (monoparasitism) of *Ancylostoma caninum* and March (20.00%) in *Ancylostoma caninum* occurrence. The lowest occurrence was recorded in (16.6%), followed by June-July (4.04%) then October and September, gave a high prevalence of 33.33% in patterns in *Diphylidium caninum* infections. *Toxocara canis* infection was age dependent showing a decreasing prevalence with age of host. *Toxocara canis*, *Ancylostoma caninum* and *Diphylidium caninum* were the zoonotic gastrointestinal helminthes prevalent in dogs in the study area. This study provides a baseline data and evidence that Zoonotic infection is prevalent in dogs in the study area, therefore, the need to educate the residents of Jos Metropolis on the danger of close association or companion with their dogs.

**Keywords:** Zoonotic gastrointestinal parasites, Dogs, *Toxocara canis*, *Ancylostoma caninum*, *Diphylidium caninum*

### INTRODUCTION

Zoonotic infections can be defined as infections of animals that are naturally transmissible to humans [1]. In Nigeria, occurrences of these zoonotic parasitic infections have been reported widely in dogs with differences in prevalence depending on the geographical location [2-5], in this regard, dogs are considered as a public health concern, as they may harbor various pathogens and often spread the infection to humans through their association as companion or domestic animals [6]. The transmission of zoonotic agents could be through indirect contact with animal secretions and excretions, infected water and food and through direct contact with the animal [7]. Intestinal helminthes are among the most common pathogenic agents encountered in dogs, especially in newly whelped or neonates and cause pathologies in the intestine

[8]. Some of the most important and well-known human zoonotic helminthic diseases are, toxocariasis or visceral larva migrans, ancylostomitidosis or cutaneous larva migrans, tungiasis, hydatid disease as well as emerging and re-emerging infections such as cryptosporidiosis and giardiasis

**Corresponding author:** Gloria Pisha Karaye, Department of Veterinary Parasitology and Entomology, University of Jos, Plateau State, Nigeria, Tel: +234 (0) 8060926642; E-mail: Pishluv2@yahoo.com

**Citation:** Karaye GP, Kaze PD, Akinsola OM, Wamtas FI, Kogi AC et al. (2021) Gastrointestinal Health Parasites of Domestic Dogs in Jos North, Plateau State Nigeria: A Fecal Examination Study. J Vet Marine Sci, 3(1): 136-142.

**Copyright:** ©2021 Karaye GP, Kaze PD, Akinsola OM, Wamtas FI, Kogi AC et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

[9-11], species of nematodes like, (trichinellosis), cestodes (cysticercosis, echinococcosis) and trematodes (schistosomiasis) [12]. Others include intestinal capillariasis, anisakidosis, eosinophilic enteritis, oesophagostomiasis and gnathostomiasis [13].

*Toxocara canis* is the causative agent of toxocariasis and its one of the most common gastrointestinal helminthes living in the intestines of domestic and stray dogs [3]. It has been reported in nearly all parts of the world with prevalence of up to 100% in some population of puppies [14]. Transmission of toxocariasis occurs in different ways depending upon the age and management of dogs. Adult dogs become infected with *T. canis* through ingestion of infective eggs or infective larvae in tissue of paratenic hosts such as mice, birds, pigs, earthworms and others [15]. Various surveys conducted worldwide indicate that prevalence of *Toxocara* spp. infection in canid definitive hosts ranged from 86-100% in puppies and 1-45% in adult dogs [16-19].

The clinical symptoms of Toxocariasis caused by *T. cati* and *T. canis* frequently impacts young cats and dogs from birth to 1 year old, entailing respiratory signs (coughing due to pulmonary larval migration), general failure to thrive (retarded growth, emaciation, debilitated body coat and arthralgia) and intestinal disorders (alternating diarrhea and constipation, pot-belly and vomiting). Death is rare but has been reported in severe cases as a result of obstruction of the intestine or ulceration and perforation on the intestinal wall [20-22].

In Nigeria, gastrointestinal helminthes parasites of dogs are currently endemic in 20 of the 36 States [23,24]. Some of the emerging infections are due to the prevailing socio-economic conditions in Nigeria which have made it difficult for many dog owners to adequately provide food, shelter and basic health needs for their dogs. These have resulted in increased number of dogs scavenging for food on the streets and increasing the risk of human infection among the communities.

This study was carried out with the aim of providing information on the diagnosed parasitic conditions of owned dogs in the State. This is to add to the existing information as well as to the knowledge of the epidemiology of parasitic diseases of dogs in Nigeria and as such assist the government in formulating policies on the parasitic diseases of dog prevalent in the study area as well as their zoonotic implications to the persons residing in the area This study was carried out with the aim of providing information on the diagnosed parasitic conditions of owned dogs in the State. This is to add to the existing information as well as to the knowledge of the epidemiology of parasitic diseases of dogs in Nigeria and as such assist the government in formulating policies on the parasitic diseases of dog prevalent in the study area as well as their zoonotic implications to the persons residing in the area

The present study attempts to determine and provide information on the diagnosed parasitic conditions of owned dogs harboring *Toxocara canis* and other potential public health significance intestinal parasites in Jos North LGA of Plateau, Nigeria in a bid to add to the existing information as well as to the knowledge of the epidemiology of parasitic disease of dogs to the persons residing in this areas.

## MATERIALS AND METHODS

This study was carried out at the Veterinary teaching hospital of the Faculty of Veterinary Medicine, University of Jos, Plateau State, and North-Central Nigeria. The University of Jos is geographically located in Jos North Local Government Area which covers an area of 291 square kilometers with a population of 429,300 at the 2006 census [25]. Plateau State on the other hand, covers an area of 27,147 square kilometers and is one of the largest States in Nigeria. It is almost centrally located between Latitude 80°24'N and Longitude 80°32' and 100°38' east of the Greenwich meridian. The State has a high altitude ranging from approximately 1,200 to a peak of 1,829 meters above sea level. Plateau State has a near temperate climate with an annual rainfall of between 131.75 cm to 146 cm and an average annual temperature ranging between 16.3°C and 28.1°C. It records a mean relative humidity of between 46.9% and 51.3% [25,26]. The major ethnic groups on the Plateau are Anaguta, Afizere and Berom, Miango and Tarok; other settlers include, Hausa, Yoruba, Igbo, etc.

## STUDY DESIGN

A total of 321 samples obtained from different breeds of dog (Caucasian, Rottweiler, Alsatians, cross breed and Nigerian indigenous) were collected and analyzed. The dogs comprised of clinically healthy and clinically sick animals. The sampling was conducted from January 2019 to March 2020. The owners of the dogs were approached and the purpose for the sampling was explained, after which their cooperation was sought for the collection of fecal sample and questions were asked the dog owners to obtain information as regards the approximate age, sex, mode of life, breed type and disease related knowledge of the owners. This information, therefore, gave the bases for the classification of the dogs.

## FAECAL SAMPLE COLLECTIONS AND PROCESSING

Sterile forceps were inserted into the rectum of the dogs and transferred into sterile, sample bottles which were properly labeled with each dog's identification number (ID), age and sex. The fecal samples collected were first examined physically for larvae, adult worms and tapeworm eggs before transferring some quantity into the sample bottles. The fecal samples were immediately analyzed in the parasitology laboratory of the Department of Parasitology and Entomology, University of Jos, for processing and the recovery of helminth eggs. Fecal samples were processed for microscopic examination for helminth eggs by modified Kato-Katz technique [27]. Briefly, the fecal samples were

first homogenized by shaking vigorously to form a paste using a clean glass rod, were after a small portion of the sample was sieved through double-ply gauze to remove debris and rough materials. The filtrate was centrifuged at 2500 rpm for 5 min, the supernatant decanted and the tube was allowed to stand for 2 min. 41.7 mg of the sediment delivered by Kato-Katz template was taken onto a degreased glass slide and covered with a cellophane strip soaked overnight in 50% solution of glycerol-malachite green. Slides were examined for helminth eggs under a light microscope. Observed helminth ova in 41.7 mg of sieved stool were identified using known structural and morphometric features [28]. In addition to qualitative diagnosis, indirect measure of helminth intensity was obtained by counting eggs and expressed as eggs/gram of feces (EPG) by multiplying the number of eggs counted with a factor of 24 [29].

### STATISTICAL ANALYSIS

For the statistical analysis of the data, the animals were grouped by age (above 4 years, below 2 years between 2 to 3 years old), Breed type (local and exotic), month of sampling

(January to March, 2020) the overall prevalence for all parasites and the particular prevalence of each helminth were determined. Data generated were analyzed using descriptive statistics (frequency and percentage) and chi square test of SPSS 20 for windows, version 17 (Chicago, Illinois, USA) to determine the effect between categorized variables of parasite, year, age and breed in association with the occurrence of parasites.

### RESULTS

**Table 1** describes prevalence of *Toxocara canis* relative to the age of dogs presented to the university of Jos teaching hospital from January 2019 to December 2019. **Table 2** describes the prevalence of *Toxocara canis* relative to the breeds of dogs presented to the university of Jos teaching hospital from January 2019 to December 2019 and **Table 3** describes the prevalence of *Toxocara canis* relative to the months of the year presented to the university of Jos teaching hospital from January 2019 to December 2019.

**Table 1.** Prevalence of *Toxocara canis* relative to the age of dogs.

Parasites	Above 4 years	Below 2 years	2 to 3 years	Total
<i>Ancylostoma caninum</i> and <i>Toxocara canis</i>	6(19.35%)	13(41.94%)	12(38.71%)	31(9.96%)
<i>Ancylostoma caninum</i>	4(16.00%)	13(52.00%)	8(32.00%)	25(7.81%)
<i>Diphylidium caninum</i>	3(50.00%)	1(16.67%)	2(33.33%)	6(1.88%)
No Parasite seen	30(18.87%)	87(54.72%)	42(26.42%)	156(49.69%)
<i>Toxocara canis</i>	26(26.26%)	44(44.44%)	29(29.29%)	99(30.94%)

$$\chi^2 = 8.807; df = 8 P = 0.3588$$

**Table 2.** Prevalence of *Toxocara canis* relative to the breeds of dogs presented to the university of Jos teaching hospital from January 2019 to December 2019.

Parasites	Bull Mastiff	Alsations	Caucasians	Indigenous	Cross	Total
<i>Ancylostoma caninum</i> and <i>Toxocara canis</i>	7 (22.58%)	5 (16.13%)	1 (3.23%)	11 (35.48%)	7 (22.58%)	31 (9.69%)
<i>Ancylostoma caninum</i>	4 (16.00%)	6 (24.00%)	1 (4.00%)	10 (40.00%)	4 (16.00%)	25 (7.81%)
<i>Diphylidium caninum</i>	3 (50.00%)	0 (0.00%)	0 (0.00%)	1 (6.67%)	2 (33.33%)	6 (1.88%)
No Parasite seen	25 (15.72%)	31 (19.50%)	22 (13.84%)	49 (30.82%)	32 (20.133%)	159 (49.69%)
<i>Toxocara canis</i>	14 (14.14%)	25 (25.25%)	13 (13.13%)	30 (30.30%)	17 (17.17%)	99 (30.94%)

$$\chi^2 = 15.039; df = 16 P = 0.5218$$

In total, 321 fecal samples were analyzed, and results showed that the dogs were infected with one or more helminth species. Eggs of three helminth species comprising two nematodes and one cestode were identified.

As depicted in **Table 1** above, the prevalence of *Toxocara canis* was highest in dogs less than 2 years of age followed by those that are between 2 to 3 years of age, then the least prevalence was recorded in dogs that are 4 years and above. There was no significant difference ( $P > 0.05$ ) in the overall

prevalence of *Toxocara canis* infection in dogs that are less than 2 years (44.44%, 54.72%, 16.67%, 52.00%, 41.94%), Dogs within the age brackets of 2 to 3 years (22.29%, 26.42%, 33.33%, 32.00%, 38.71%) and those above 4 years (26.26%, 18.87%, 50.00%, 16.00%, 9.35%) respectively.

**Table 3** describes the prevalence of *Toxocara canis* relative to the months of the year presented to the university of Jos teaching hospital from January 2019 to December 2019.

Paraasites	Jan	Feb	Mar	Apr	May	Jun	July	Aug	Sept	Oct	Nov	Dec	Total
<i>Ancylostoma caninum</i> and <i>Toxocara canis</i>	11 (35.48%)	5 (16.13%)	5 (16.13%)	3 (9.68%)	0 (0.00)	2 (6.45%)	0 (0.00%)	1 (3.23%)	0 (0.00%)	0 (0.00%)	3 (9.68%)	1 (3.23%)	31 (9.69%)
<i>Ancylostoma caninum</i>	3 (12.00%)	5 (20.00%)	5 (20.00%)	2 (8.00%)	2 (8.00%)	1 (4.00%)	0 (0.00%)	1 (4.00%)	3 (12.00%)	1 (4.00%)	0(0.00%)	2(8.00%)	25 (7.81%)
<i>Dipylidium caninum</i>	2 (33.33%)	0 (0.00%)	0 (0.00%)	1 (16.67%)	1 (16.67%)	0 (0.00%)	0 (0.00%)	0 (0.00%)	0 (0.00%)	0 (0.00%)	2 (33.33%)	0(0.00%)	6(1.88%)
No parasite seen	44 (27.67%)	27 (16.98%)	12 (7.55%)	13 (8.18%)	10 (6.29%)	6 (3.77%)	6 (3.77%)	10 (6.29%)	9 (5.66%)	5 (3.14%)	9 (5.66%)	7(4.40%)	159 (49.69%)
<i>Toxocara canis</i>	30 (30.30%)	9 (9.09%)	14 (14.14%)	11 (11.11%)	2 (2.02%)	4 (4.04%)	4 (4.04%)	6 (6.06%)	1 (1.01%)	8 (8.08%)	7(7.07%)	3(3.03%)	99 (30.94%)

$$X^2 = 49.658; p = 0.4070; df = 48$$

The prevalence of *Toxocara canis* was significantly higher in (Nigerian Indigenous dogs ) breed (30.30%, 30.82%, 16.67%, 40.00% and 35.48%) than the exotic breeds Bullmastif (14.14%, 15.72%, 50.00%, 16.00%, 22.50%), Alsatian

(25.25%, 19.50%, 0.00%, 24.00%, 16.13%), Caucassian (13.13%, 13.84%, 0.00%, 4.00%, 3.23%) and least prevalence was recorded in the Cross breed (17.17%, 20.13%, 33.33%, 16.00% and 22.58%) respectively. Although, there was no significant difference recorded in the overall groups ( $P>0.05$ ) (Table 2).

The prevalence of *Toxocara canis* infection was higher in January (30.30%, 27.60%, 33.33%, 12.00%, 35.48%) followed by March (11.11%, 18.18%, 16.67%, 8.00%, 9.68%), April (11.11%, 18.18%, 16.67%, 8.00%, 9.68%), February (9.09%, 16.98%, 0.00%, 20.00%, 16.13%), August (6.06%, 6.29%, 0.00%, 4.00%, 3.23%), November (8.08%, 7.40%, 33.33%, 8.00%, 9.68%), December (7.07%, 4.40%, 0.00%, 8.00%, 3.23%). The least prevalence was recorded in the months of May (2.02%, 6.29%, 0.00%, 8.00%, and 0.00%), June (4.04%, 3.77%, 6.00%, 4.00%, and 6.45%) and July (0.00%, 4.00%, 0.00%, 3.77% and 4.04%) respectively. Based on the trend of infection within the months, there was no significant difference recorded from January to December (Table 3).

## DISCUSSION

This study provides the first assessment of the common zoonotic helminthes namely, *Toxocara canis*, hookworm (*Ancylostoma caninum*) and *Dipylidium caninum* parasites in dogs presented at the Veterinary teaching hospital Jos, Jos North Plateau State. These helminthes, as shown in Table 1, are more prevalent among young puppies of less than two years of age. The observed zoonotic parasites in this study, has already been reported in different studies and at different locations within Jos [30,4,5] and in other geographical regions in Nigeria [31,11,32,24]. The high prevalence of ascarid infections in puppies is in accordance with the transmission pattern of the parasite, which is mainly by transplacental, percutaneous and transmammmary routes; acquired age-dependent immunity may be caused by repeated exposure which could have been the reason why the older dogs had less infection [33,34]. A similar study [35] however reports that parasite species which are not transmitted to dogs at early age, do not elicit a specific immune response and so there is an increased infection rate in older dogs.

Sopora species, *Ancylostoma caninum*, *Taenia ovis* and *Toxocara canis* were the gastrointestinal parasites detected in dogs from our study with *Ancylostoma caninum* and *Toxocara canis* been the most prevalent. Similar to our finding, *Ancylostoma caninum* and *Toxocara canis* has been reported to be the most prevalent gastrointestinal parasites of dogs in Nigeria [36-38].

Polyparasitism (*Toxocara canis* and *Ancylostoma caninum*) with more than one parasite species of dogs was also observed in this study and this is similar to other reports on the occurrence of these two parasites as the most prevalent helminthes parasite in Nigerian dogs [36-38,5]. Also, reported that common occurrence of co-infections by intestinal

helminthes possible and might be as a result of a heavy environmental contamination, suggesting that animals (and people in the case of zoonotic parasites) sharing the same habitat are at high risk of infection. The presence of infective eggs or larvae in the environment has a crucial role among the different routes of transmission of dog intestinal nematodes in both humans and animals. In fact, human beings become infected by canine *Toxocara* species and *Ancylostoma* spp. most frequently through contaminated soil [39].

*Toxocara canis* is one of the helminth observed in this study with a high prevalence rate. *Toxocara canis* is the causative agent of toxocariasis and its one of the most common gastrointestinal helminthes living in the intestines of domestic and stray dogs [3] and it is still a seriously notifiable public health issue, particularly due to its intricate transmission routes [40]. It has been reported in nearly all parts of the world with prevalence of up to 100% in some population of puppies [14]. Transmission of toxocariasis occurs in different ways depending upon the age and management of dogs. Adult dogs become infected with *T. canis* through ingestion of infective eggs or infective larvae in tissue of paratenic hosts such as mice, birds, pigs, earthworms and others [15].

*Dipylidium caninum* is a zoonotic tapeworm that was recovered among the dog gastrointestinal parasites in all the variables. Dogs, cats and wild carnivores are the definitive hosts of these tapeworms, although, occasionally man can be an accidental host. [41]. The presence of this parasite in the study area is of public health importance, particularly to children. Children have been reported to be more likely infected with dipylidiasis than adults [42]. Owing to their adventurous nature as children love to put objects picked on the floor directly into their mouth for tasting, eating soil, or fruits from the floor not washed and being in the vicinity of dogs and cats, they are considered as a substantial risk group regarding toxocariasis [43,20].

This is of *Toxocara canis* and co infection of *Toxocara canis* and the disparity in prevalence of intestinal parasite observed among local, cross and exotic breeds of dogs in this study may be associated with density and species composition of parasites observed. It may also reflect the degree of environmental contamination and inequalities in the health care services between urban and rural areas as well as differences in handling and health care by the dog owners resulting in the reduction of intestinal parasite burden. It is also worthy of note that exotic and cross breed dogs which usually serve as pets or security dogs, are usually acquired at high costs and their keepers invest in deworming and other routine treatments. Conversely, the prevalence of gastric helminthes in local dogs could be attributed to the lack of inadequate care they receive as compared to the exotic ones [35,5]; these dogs are usually allowed to roam about to fend for themselves and treatments are often administered only when the owner observes some changes in the animal behavior or helminth segments in its feces.

Meanwhile, investigation on the monthly distribution of gastrointestinal helminths revealed that all the cestodes and nematodes species were present all year round among all the observed variables. A steady increase in number was observed from January-April; which coincided with period of extreme dry season (early and late). However, a gradual decline in parasite abundance was noticed from May-August, a period characterized by high rainfall similar findings were reported in North-eastern Nigeria [44], North-central Nigeria [5], as well as Ibadan. It has been established that during the dry season, we presume this is the period some of the climatic factors are optimal for the proliferation of the etiology agent; hence favoring helminth nematodes multiplication and spread. Besides, the environmental stress experienced during the dry season might exert a negative influence on the immune system of the host, thereby increasing the likelihood for infection to occur [45,46].

Since no previous data were available regarding the treatment history or signalment of the dogs from which fecal samples were examined, additional factors could be influencing the data, that is, it is possible that the percentage of dogs and puppies in presented to the clinic is not the true representation of the population of dogs in the study area and that some animals were coming as follow up cases thereby influencing prevalence. It is also possible that seasonal usage of dewormers for the prevention in some parts of the Jos, could be lowering prevalence during the wet months of the year [47,48].

Season was significantly associated with the prevalence of helminthoses, tick infestation, lice infestation and myiasis in our study. Higher prevalence of these parasitic condition occurred in the wet season compared to the dry season with exception of helminthoses reported a significantly higher prevalence of endoparasitic and ectoparasitic conditions in dogs during the wet season than the dry season. Climatic seasons are determined by the amount of rainfall, mean temperature, relative humidity, solar radiation and wind strength, which are important indices in the epidemiology of parasitic diseases in dogs as these either favors or hinders the survival of parasites and its vectors, the transmission of parasitic diseases and the development of diseases in the host [49].

## CONCLUSION

In conclusion, this study has revealed that the three gastrointestinal helminth parasites identified i.e *Toxocara canis*, *Ancylostoma caninum* and *Dipylidium caninum* are zoonotic parasites which are common in Jos, Jos North LGA of Plateau State and all classes of dogs are affected, with high prevalence which constitutes a public health problem in the study area. Hence, interventive measures are necessary to reduce the risk of transmission of parasites from dogs to humans and that public education on the proper care of dogs including veterinary care, personal hygiene by dog owners

and handlers, are recommended and prevention of zoonotic parasitic diseases are of great importance in the study area.

## REFERENCES

1. Goldsmid JM (2005) Zoonotic infections - An overview, Chapter 14, pp: 14.1-14.14. Available online at: <https://www.tropmed.org/wp-content/uploads/2018/05/chapter14.pdf>
2. Magaji AA, Mohammed MN, Saulawa MA, Salihu MD (2012) Survey of zoonotic gastrointestinal parasites of dogs (*Canis familiaris*) slaughtered at Zuru area, Kebbi State, Nigeria. *Sci J Vet Adv* 1: 132-136.
3. Ogbaje CI, Ademola IO (2014) Prevalence of zoonotic gastrointestinal parasite burden of local dogs in Zaria, Northern Nigeria: Implication for human health. *Int J One Health* 1: 32-36.
4. Chanding AY, Umar YA, Tenshak TF, Ibrahim S (2018) Prevalence study of the gastrointestinal helminth in dogs (*Canis familiaris*) slaughtered in selected Abattoirs in Plateau State. *Niger Open Sci J* 3.
5. Karaye GP, Fadunsin SDO, Dogo DA (2018). Diversity of gastrointestinal parasites affecting some domestic animals in Plateau State, North Central Nigeria. *Sci World J* 13: 81-86.
6. Berrett AN, Erickson LD, Gale SD, Stone A, Brown BL, Hedges DW (2017) *Toxocara* seroprevalence and associated risk factors in the United States. *Am J Trop Med Hyg* 97: 1846-1850.
7. Lappin MR (2002) Pet ownership by immune compromised people, Bayer zoonosis symposium, North American Conference. 24: 16-25.
8. Blagburn BL, Lindsay DS, Vaughan JL, Rippey NS, Wright JC (1996) Prevalence of canine parasites based on fecal flotation. *Compend Contin Educ Vet* 18: 483-509.
9. Heukelbach J, Wilcke T, Meier A, Moura RC, Feldmeier H (2003) A longitudinal study on cutaneous larva migrans in an impoverished Brazilian township. *Travel Med Infect Dis* 1: 213-218.
10. Akao N, Ohta N (2007) Toxocariasis in Japan. *Parasitol Int* 56: 87-93.
11. Ugbomoiko US, Ariza L, Heukelbach J (2008) Parasites of importance for human health in Nigerian dogs: High prevalence and limited knowledge of pet owners. *BMC Vet Res* 4: 49.

12. Deplazes P, Knapen FV, Schweiger A, Overgaauw PA (2011) Role of pet dogs and cats in the transmission of Helminthic zoonoses in Europe, with a focus on Echinococcosis and Toxocariasis. *Vet Parasitol* 24: 41-53.
13. Stojeeviae D, Susiae V, Lueinger S (2010) Contamination of soil and sand with parasite elements as a risk factor for human health in public parks and playground in Pula Croatia. *Vet Arh* 80: 733-742.
14. Overgaauw PA (1997) Prevalence of intestinal nematodes of dogs and cats in the Netherlands. *Vet Q* 19: 14-17.
15. Maqbool A, Raza SH, Hayat CK, Hafiq M (1998) Prevalence and chemotherapy of toxocariasis in the dog in Faisalabad (Punjab). *Pak Vet Arch* 68: 121-125.
16. Dai RS, Li ZY, Li F, Liu DX, Liu W (2009) Severe infection of adult dogs with helminths in Hunan Province, China poses significant public health concerns. *Vet Parasitol* 160: 348-350.
17. Soriano SV, Pierangeli NB, Roccia I, Bergagna HF, Lazzarini LE (2010) A wide diversity of zoonotic intestinal parasites infects urban and rural dogs in Neuquen, Patagonia, Argentina. *Vet Parasitol* 167: 81-85.
18. Overgaauw PA, Knapen FV (2013) Veterinary and public health aspects of *Toxocara* spp. *Vet Parasitol* 193: 398-403.
19. Schär F, Inpankaew T, Traub RJ, Khieu V, Dalsgaard A, et al. (2014) The prevalence and diversity of intestinal parasitic infections in humans and domestic animals in a rural Cambodian village. *Parasitol Int* 63: 597-603.
20. Macpherson CN (2013) The epidemiology and public health importance of toxocariasis: A zoonosis of global importance. *Int J Parasitol* 43: 999-1008.
21. Lee RM, Moore LB, Bottazzi ME, Hotez PJ (2014) Toxocariasis in North America: A systematic review. *PLoS Neglect Trop D* 8: e3116.
22. Lötsch F, Vingerling R, Spijker R, Grobusch MP (2017) Toxocariasis in humans in Africa: A systematic review. *Travel Med Infect Dis* 20: 15-25.
23. Uwemedino E, Akinola O, Alexios KVD, Eniola A, Franca O, Sunday I (2014) Bayesian geostatistical model-based estimates of geospatial distribution of soil transmitted Helminthiasis and Albendazole Treatment Requirements in Nigeria. 13th International Congress of Parasitology, August 10th-15th Mexico.
24. Christopher IO, Raphael AO, Ikwe AA (2015) Zoonotic gastrointestinal parasite burden of local dogs in Zaria Northern Nigeria. Implication for human health. *Int J One Health* 1: 32-36.
25. National Bureau of Statistics (NBS) (2012) Annual Abstract of Statistics. Federal Republic of Nigeria.
26. Bolajoko M, Ahmed MS, Okewale PA, Kumbish P, Mohammed M, et al. (2016) Prevalence and demographic distribution of canine rabies in Plateau State, Nigeria 2004-2009. *Bull Anim Health Prod Afr* 64: 127-136.
27. Forrester JE, Scott ME (1990) Measurement of *Ascaris lumbricoides* infection intensity and dynamic of expulsion following treatment with mebendazole. *Parasitol* 100: 303-308.
28. Bowman DD (1999) *Georgis' parasitology for veterinarians*. London, WB Saunders.
29. Katz N, Chaves A, Pellegrino J (1972) A simple device for quantitative stool thick-smear technique in *Schistosomiasis mansoni*. *Rev Inst Med Trop Sao Paulo* 14: 397-400.
30. Pam VA, Ogbu KT, Akinyera AO, Gullek JT, Okoro J (2015) Investigation on the prevalence of gastrointestinal parasites in local and exotic dogs in Jos South Local Government Area of Plateau State, Nigeria. *Int Res J Public Health* 2: 55-60.
31. Sowemimo O, Asaolu SO (2008) Epidemiology of intestinal helminth parasites of dogs in Ibadan, Nigeria. *J Helminthol* 82: 89-93.
32. Ademola IO, Odeniran P (2013) Prevalence of zoonotic gastrointestinal Helminth in dogs and knowledge of risk of infection by dog owners in Ibadan, Nigeria. *Niger Vet J* 34: 851-858.
33. Pereckiene A, Kaziūnaite V, Vysniauskas A, Petkevicius S, Malakauskas A (2007) A comparison of modifications of the McMaster method for the enumeration of *Ascaris suum* eggs in pig fecal samples. *Vet Parasitol* 149: 111-116.
34. Othman RA (2011) Prevalence of *Toxocara canis* in dogs, North West Bank of Palestine. *Korean J Parasitol* 49: 181-182.
35. Sowemimo O, Ayanniyi OA (2017) Gastrointestinal helminth parasites of domestic dogs in Ilesa, Osun State, Nigeria: A fecal examination survey study. *J Bacteriol Parasitol* 8: 3.
36. Adamu NB, Adamu JY, Salisu L (2012) Prevalence of ecto, endo-and haemoparasites in slaughtered dogs in Maiduguri, Nigeria. *J Vet Med* 4: 178-182.

37. Mustapha FB, Balami SB, Malgwi SA, Adamu SG, Wakil Y (2016) Prevalence of gastrointestinal tract parasites of hunting dogs in Maiduguri, Borno, Nigeria. *IOSR J Agric Vet Sci* 9: 39-42.
38. Idika IK, Onuorah EC, Obi CF, Umeakuana PU, Nwosu CO, et al. (2017) Prevalence of gastrointestinal helminth infection in dogs in Enugu State, South Eastern Nigeria. *Parasite Epidemiol Control* 2: 97-104).
39. Traversa D (2012) Pet roundworms and hookworms: A continuing need for global warming. *Parasitol Vector* 5: 91.
40. Elefant GR, Hirata CE, Yamamoto JH, Ferreira MU (2010) Human toxocariasis: Diagnosis, worldwide seroprevalences and clinical expression of the systemic and ocular forms. *Ann Trop Med Parasitol* 104: 3- 23.
41. Adam AA, Abu OAR, Abdel SMN, Abdel MNI (2006) Cutaneous myiasis due to *Dermatobia hominis*: A case report eastern Sudan. *Sudan J Med Sci* 1: 147-148.
42. Molina CP, Ogburn J, Adegboyega P (2003) Infection by *Dipylidium caninum* in an infant. *Arch Pathol Lab Med* 127: 157-159.
43. Zibaei M, Sadjjadi SM, Sarkari B (2007) Prevalence of *Toxocara cati* and other intestinal helminths in stray cats in shiraz, Iran. *Trop Biomed* 24: 39-43.
44. Mohammed K, Biu A, Ahmed MI, Charles A (2014) Prevalence and seasonal abundance of ticks on dogs and the role of *Rhipicephalus sanguineus* in transmitting *Babesia* species in Maidugiri, North-Eastern Nigeria. *Vet World* 7: 119-124.
45. Anene BM, Nnaji TO, Chime AB (1996) Intestinal parasitic infections of dogs in the Nsukka area of Enugu state, Nigeria. *Preview Vet Med* 27: 89-94.
46. Diakou A, Dicesare A, Acettura MP, Baros L, Iono R, et al. (2017) Intestinal parasites and vector borne pathogens in stray and free roaming cats living in continental and insular Greece. *PLoS Negl Trop Dis* 11: e0005335.
47. Ontanarrosa MF, Vezzani D, Basabe J, Eiras DF (2006) An epidemiological study of gastrointestinal parasites of dogs from Southern Greater Buenos Aires (Argentina): Age, gender, breed, mixed infections, and seasonal and spatial patterns. *Vet Parasitol* 136: 283-295.
48. Omudu EA, Amuta EU (2007) Parasitology and urban livestock farming in Nigeria: Prevalence of ova in faecal and soil samples and animal ectoparasites in Makurdi. *J Sci Afr Vet Assoc* 78: 40-45.
49. Senlik B, Cirak VY, Karabacak A (2006) Intestinal nematode infections in Turkish military dogs with special reference to *Toxocara canis*. *J Helminthol* 80: 299-303.