

## Investigating Antimicrobial Effects of *Tecomella undulata* Ethanolic Extract on Antibiotic Resistant *Acinetobacter baumannii*

Khadije Saravani<sup>1</sup> and Fereshteh Javadian<sup>2\*</sup>

<sup>1</sup>Faculty of Medicine, Zabol University of Medical Sciences, Zabol, Iran

<sup>2</sup>Zabol Medicinal Plant Research Center, Zabol University of Medical Sciences, Zabol, Iran.

Received July 23, 2019; Accepted July 31, 2019; Published April 08, 2020

### ABSTRACT

Considering the increasing resistance of bacteria to antibiotics and the presence of antibacterial agents in plants, in this study, the antimicrobial activity of *Tecomella undulata* ethanolic extract on antibiotic resistance *Acinetobacter baumannii* has been studied.

**Materials and methods:** The leaves of *Tecomella undulata* were collected from Saravan city and extracted by rotary machine. *Acinetobacter bomanii* strains were collected from urine specimens of Imam Khomeini and Ali ibn Abi Talib Hospitals. Minimum inhibitory concentration and minimum bactericidal concentration were determined by micro dilution method.

**Results:** The results of this study showed that the resistance of the strains was to amoxiclavanic (10%), ampicillin (20%), gentamicin (0%), ceftazidime (0%) and nitromicin (0%) antibiotics.

The results of this study showed that the lowest inhibitory concentration of *Tecomella undulata* is 0.62 mg/ml, which inhibits 6 strains in this concentration, while the highest inhibitory concentration is 5 mg/ml, which inhibited 3 strains in this concentration.

**Discussion:** By considering the results, obtained and increasing resistance of bacteria to chemical antibiotics, it is suggested that bacterial compositions of this plant can be used to treat bacteria.

**Keywords:** *Tecomella undulata*, Antimicrobial activity, *Acinetobacter baumannii*

### INTRODUCTION

*Acinetobacter baumannii* is one of the most important pathogens in health centers that cause many infections including bacteremia, pneumonia, meningitis, urinary tract infections and ulcers. The ability to survive under various environmental conditions has made this pathogen one of the most common causes of infection in health centers [1].

*Tecomella undulata* is an Antarctic pink or embroidered pomegranate, an almost evergreen tree that runs in the southern regions of the country such as Bushehr, Fars and Hormozgan.

In addition, its distribution in Afghanistan, West Pakistan and southeastern Arabia has been recorded. Due to medicinal properties, this plant has been considered as a good treatment [2]. Flavonoid compounds, phytosterol, flavonol, fatty acids and terpenoses have been identified in various parts of the plant [3]. It has anti-inflammatory, antimicrobial and anti-oxidant activity [3,4].

This plant is useful in draining urine and enlarging the spleen. The skin of the young shoots of the plant is used for

the treatment of syphilis [5]. The purpose of this study was to evaluate the antimicrobial activity of the ethanolic extract of *Tecomella undulata* on the antibiotic-resistant *Acinetobacter bomanii* in Zabol.

### MATERIALS AND METHODS

In this study 20 isolates of *Acinetobacter baumannii* from infected patients in Imam Khomeini and Ali ibn Abi Talib Hospitals in Zabol were investigated.

**Corresponding author:** Fereshteh Javadian, Zabol Medicinal Plant Research Center, Zabol University of Medical Sciences, Zabol, Iran, E-mail: fereshteh.javadian@yahoo.com

**Citation:** Saravani K & Javadian F. (2020) Investigating Antimicrobial Effects of *Tecomella undulata* Ethanolic Extract on Antibiotic Resistant *Acinetobacter baumannii*. J Genet Cell Biol, 3(1): 140-144.

**Copyright:** ©2020 Saravani K & Javadian F. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Laboratory procedures

The clinical specimens were cultured on the McConky Agar and Blood Agar medium then plates were incubated at 37°C for 24-24 h. An oxidase test was performed in case of growth after gram staining and observation of cocci and gram negative diplococci. In the next step, by using biochemical tests, cultured on McConky agar and incubated at 37°C and 42°C, then citrate and moving test were performed on the media containing glucose.

### Determination of antibiotic susceptibility

Determination of susceptibility was done by standard Disc diffusion agar. For testing, bacterial colonies, 0.5 µM MacFarland suspensions were prepared and well-spread over the Muller Hinton Agar medium. Then Antibiotic discs were placed at standard spacing. After 24 h incubation at 37°C, the non-growth diameter for each antibiotic was measured. The results were recorded for each antibiotic according to the relevant instructions as sensitive, intermediate and resistant.

### Preparation of ethanolic extract

*Tecomella undulata* collected from Saravan city and dried. To prepare the ethanolic extract, 10 g of dried powder were placed inside half-liter erlenmeyer flask containing 100 ml of 96% ethanol (to prepare the ethanolic extract). The contents of the erlenmeyer flask were mixed at room temperature for 24 h by shaker machine (Iran) at 130 rpm and then filtered with Whatman No. 2 paper. Solvent separation from the extract was performed by a rotary machine (Heidolph-Germany) with the aid of a vacuum pump (vacuum distillation). The extracts

were weighed and then solved in DMSO solvent. The extract was stored in a refrigerator until use in antimicrobial experiments at 4°C.

### Determination of susceptibility of bacterial strains to different extracts of pomegranate plant

Determination of susceptibility of bacterial strains to plant extracts was performed using a dilution method in well. Six wells were created in a solid culture medium and 100 µl of each well was added to the nutrient medium of Muller Hinton (MHB). Then, to the first well, 100 µl of diluted solution of the extracts of plants was added and after mixing 100 µl of the first well, added to the second well, and this was done until the last well. From the final well, 100 µl of the medium was extracted and 10 µl of the microbial suspension containing 10<sup>7</sup> µg/ml which was equal to 0.5 McFarland added and incubated at 37°C for 24 h. The first well that was prevented bacterial growth after placing in the incubator was considered as the minimum inhibitory concentration. In order to ensure, 10 µl from transparent wells were transferred to the Muller Hinton Agar medium and after 24 h the first concentration that could eliminate 99.9% of the bacteria was considered as the minimum bactericidal concentration.

### RESULTS

The results of this study showed that the strains were resistant to amoxiclavanic antibiotics (10%), ampicillin (20%), gentamicin (0%), ceftazidime (0%) and erythromycin (0%) (**Table 1**).

**Table 1.** Antibiotic pattern.

Antibiotics	Resistant (%)	Intermediate (%)	Sensitive (%)
AMC	1 (10%)	1 (10%)	8 (80%)
AM	2 (20%)	3 (30%)	5 (50%)
GM	0 (0%)	0 (0%)	0 (100%)
CZ	0 (0%)	0 (0%)	0 (100%)
AZM	0 (0%)	0 (0%)	0 (100%)

The results of this study showed that the lowest inhibitory concentration of *Tecomella undulata* was 0.62 mg/ml, of which 6 strains were inhibited at this concentration, while the highest inhibitory concentration was 5 mg/ml which three strains have been inhibited in this concentration. The highest bactericidal concentration was 10 mg/ml, which 2 strains were eliminated at this concentration, while the lowest bactericidal concentration was 1.25 mg/ml (**Table 2**).

Table 2. MIC and MBC extract plant.

Strain bacteria	MIC	MBC
1	2/5	5
2	5	10
3	2/5	5
4	0/62	1/25
5	1/25	2/5
6	5	10
7	2/5	5
8	0/62	1/25
9	0/62	1/25
10	0/62	1/25
11	2/5	5
12	0/62	1/25
13	1/25	2/5
14	0/62	1/25
15	2/5	5
16	1/25	2/5
17	5	5
18	2/5	5
19	1/25	2/5
20	2/5	5

## DISCUSSION

The results of this study showed that the strains were resistant to amoxiclalanic antibiotics (10%), ampicillin (20%), gentamicin (0%), ceftazidime (0%) and nitromicin (0%) [6].

The results of this study showed that the highest resistance was to ceftriaxone, ciprofloxacin and cefotaxime, which was 99%, observed in Angoti et al. [7], who investigated the drug resistance of *Acinetobacter baumannii* strains in Imam Reza Hospital. The percentage of isolates resistance to ampicemin, amikacin and ciprofloxacin was 73.3%, 38.3% and 93.3% in the E-test method, respectively.

The results of Rastu and Bad [8], who investigated the *Acinetobacter baumannii* resistance pattern in Shariati Hospital in Tehran, showed that the highest sensitivity was to ciprofloxacin (91%), cotrimoxazole (57.5%) and the highest resistance rate was to ceftriaxone (98.4%).

In the study of Nah et al. [9], who evaluated the resistance level of *Acinetobacter baumannii* strains in Tehran city, the results showed that all isolates of *Acinetobacter baumannii* were resistant to ceftizoxime, cefoprazone, ceftazidime, ticarcilin, clavulanic acid, cefotaxime, aztreonam, Moropenem, cefixim, ceftriaxone, carbenicillin and ticarcylin, but all isolates were sensitive to cholestin.

The results of Ahmadikiya et al. [10], who investigated antibiotic resistance of *Acinetobacter baumannii* in Kerman, showed a resistance rate to antibiotics such as cefotaxime (100%), ceftazidime (98.9%), cefipime (100%), aztreonam (98.9%), ampicemin (97.9%), meropenem (97.9%), gentamicin (96.8%), amikacin (98.9%), ciprofloxacin (97.9%), ciprofloxacin (97.9%) and tetracycline (90.5%).

In the study of Simhon et al. [11], Sensitivity to ampicemin was 98.1% in 1990 but reduced to 64.1% in 2000 and the sensitivity to ciprofloxacin decreased from 50.5% to 13.1%. In a study by Boromand et al. [12] in Tehran, 53.4% of the

samples were resistant to ciprofloxacin and 24.6% resistant to ampicemi.

In the study by Henwood et al. [13], 46% of the isolates were resistant to ciprofloxacin and 2% of the samples were resistant to ampicemin.

The results of Zhao study [14], who investigated the resistant pattern of *Acinetobacter baumannii* showed a resistance rate to ampicillin (78.5%), cefazolin (78.5%), imipenem (92.3%), gentamicin (87.7%) and ampicillin resistance, Ceftazidime (92.3%), aztreonam (92.2%), ciprofloxacin (98.5%) and tobramycin (81.5%).

The results of Rahbar et al. [15] study on prevalence of antibiotic resistant showed that resistance to ceftriaxone (90.9%), piperacillin (90.9%), ceftazidime (84.1%), amikacin (2/85%) and ciprofloxacin (90.9%).

In the study of Uwingabiye et al. [16], resistance to antibiotics such as ciprofloxacin, ceftazidime, piperacillin-tazobactam, imipenem, amikacin, tobramycin, dabylmezine, rifampin, colistin were 87%, 86%, 79%, 76%, 52%, 43%, 33%, 32% and 1.7%, respectively.

In recent decades, the research priority has fallen down to make new and effective drugs; this is despite the fact that the world faces pathogens with drug resistance. Another concern in this regard is the cost of treating drug-resistant infections due to the higher cost of new drugs and the long time treatment of antibiotic-resistant infections than susceptible bacterial infection, which doubles the importance of finding a new method for treatment [17].

In the study of Abhishek et al. [18], the minimum inhibitory concentration of methanol extract against *B. subtilis*, *E. fecalis*, *E. coli*, *K. pneumonia*, *M. luteus*, *P. vulgaris* and *P. aeruginosa* was equal to 4-0.01-0.1-2.0 mg/ml, respectively. In Thanawala and Jolly [19], Inhibition diameter of Acetonic extract of *Tecomella undulata* was compared to *Bacillus subtilis* (17 mm) and *Staphylococcus aureus* (10 mm), while the inhibitory diameter of alcoholic extract of *Tecomella undulata* against the *Escherichia coli* was 9 mm.

## CONCLUSION

Considering the obtained results and the increasing resistance of bacteria to chemical antibiotics, it is suggested that, to conduct more studies on antibacterial compounds of this plant in treatment of bacterial infections.

## REFERENCES

- Amudhan SM, Sekar U, Arunagiri K, Sekar B (2011) OXA beta-lactamase-mediated carbapenem resistance in *Acinetobacter baumannii*. Indian J Med Microbiol 29: 269-274.
- Anonymous (1982) The wealth of India. Raw Materials, Tecoma: CSIR New Delhi 10: 136-139.
- Jain M, Kapadia R, Jadeja RN, Thounaojam MC, Devkar RV, et al. (2012) Traditional uses, phytochemistry and pharmacology of *Tecomella undulate* - A review. Asian Pac J Trop Biomed 2: S1918-S1923.
- Laghari AQ, Memon S, Nelofar A, Laghari AH (2013) *Tecomella undulata* G. Don: A rich source of flavonoids. Ind Crops Prod 43: 213-217.
- Khatri A, Garg A, Agrawal SS (2009) Evaluation of hepatoprotective activity of aerial parts of *Tephrosia purpurea* L. and stem bark of *Tecomella undulate*. J. Ethnopharmacol 122: 1-5.
- Chopra RN, Nayar SL, Chopra IC (1956) Glossary of Indian Medical Plants. 1<sup>st</sup> Edn. C.S.I.R: New Delhi 21: 240.
- Angoti G, Godarzi H, Besharat M, Hajizadeh M, Zarringhalam Moghaddam M (2014) Evaluation of antibiotic resistance of clinical *Acinetobacter baumannii* isolated of Tabriz Hospital by disk diffusion and MIC methods. Res Med 38: 106-110.
- Rastu A, Bad Y (2009) Frequency of *Acinetobacter baumannii* isolated from blood culture in a hospital laboratory Shariati Tehran. J Paramed Tehran Univ Med Sci 3: 70-75.
- Nah SF, Ranjbar R, Ghasemi A, Pakzad A, Zaemi Yazdi, et al. (2006) Evaluation of drug resistance of *Acinetobacter baumannii* and other *Acinetobacter* species isolated from three hospitals in Tehran. Sci J Elam Univ Med Sci 3: 14.
- Ahmadiqiya F, Mosadegh A, Moradi M, Hossieni-Nave H (2017) Antimicrobial resistance patterns and frequency of extended-spectrum beta-lactamase genes among *Acinetobacter baumannii*. JBUMS 19: 28-34.
- Simhon A, Rahav G, Shazberg G, Block C, Bercovier H, et al. (2001) *Acinetobacter baumannii* at a Tertiary-Care Teaching Hospital in Jerusalem. J Clin Microbiol 39: 389-391.
- Boromand M, Akyani M, Sheikhvatan R, Hekmat Yazdi S, Saboorian R, et al. (2009) Evaluation of antimicrobial resistance of *Acinetobacter baumannii* to imipenem, ciprofloxacin and ceftazidim using E test. Iran J Publ Health 2: 130-133.
- Henwood C, Gatward T, Warner M, James D, Stockolale M, et al. (2002) Antibiotic resistance among clinical isolates of *Acinetobacter* in the UK and *in vitro* evaluation of Tigecycline (GAR-936). Antimicrob Chemother 49: 479-487.
- Zhao SY, Jiang DY, Xu PC, Zhang YK, Shi HF, et al. (2015) An investigation of drug-resistant *Acinetobacter*

- baumannii* infections in a comprehensive hospital of East China. Ann Clin Microbiol Antimicrob 14: 7.
15. Rahbar M, Mehrgan H, Aliakbari NH (2010) Prevalence of antibiotic-resistant *Acinetobacter baumannii* in a 1000-bed Tertiary Care Hospital in Tehran, Iran. Indian J Pathol Microbiol 53: 290-293.
  16. Uwingabiye J, Frikh M, Lemnouer A, Bssaibis F, Belefquih B, et al. (2016) Acinetobacter infections prevalence and frequency of the antibiotics resistance: Comparative study of intensive care units versus other hospital units. Pan Afr Med J 23: 191.
  17. Overbye KM, Barrett JF (2005) Antibiotics: Where did we go wrong? Drug Discov Today 10: 45-52.
  18. Abhishek S, Ujwala P, Shivani K, Meeta B (2013) Evaluation of antibacterial activity of *Tecomella undulata* leaves crude extracts. Int Res J Biol Sci 2: 60-62.
  19. Thanawala PR, Jolly CI (1993) Phyarmacognostical phytochemical and antimicrobial studies on stem bark of *Tecomella undulate* seem. Ancient Science of Life 3-4: 414--419.